Molecular Cell, Volume 44

Supplemental Information

JNK1 Phosphorylation of Cdt1 Inhibits

Recruitment of HBO1 Histone Acetylase and Blocks

Replication Licensing in Response to Stress

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Figure S1. Related to Figure 1

Representation of HBO1 ChIP-qPCR data in percentage of total DNA

Title of each panel indicate the figure associated with these HBO1 ChIP/MCM4 origin data. See figure legend for information regarding the samples. In each panel HBO1 ChIP and IgG control data are presented for the MCM4 replication origin and the GAPDH control region.

□ IqG/MCM4 origin HBO1 ChIP/GAPDH ■IaG/GAPDH

% total 0,6 Figure 1G 0,8 %°total 0,7 0,4 0,6 Figure 1A 0,3 0,5 0,4 0,2 0,3 0,1 0,2 + 0,1 0 ----0 controlirea Aniso.DT Sorbitol controlureaanisoDTTSorbActDHU 1,4 0,9 Figure 3A 1,2 0,8 % total Figure 2C % tota 0,6 0,8 0,5 0,6 0,4 0,4 0,3 0,2 0,2 0 0,1 control - SP JNKI1038 ATMi 0 sorbitosorbitol+MG132 Sorbitol+ inhibitor control 0,9 % total 0,8 üre 3C % total Figure 3B 0,8 $0,5^{+}$ 0,6 0,4 0,4 0.3 0 0,2 0.1^{-1} 0 ctr JNK1 JNK2 control sorbitol Recoverly Sorbitol + siRNA 0.5h -1h 1 1 Figure 5D 0,9 % 69tal **DNA**Figure 3D %%total 0,6 0,6 0,5 0,4 0,4 0,3 0,2 0,2 0,1 0 0 + empty WT T29 T29 T29 Cy Empty WT T29 Ori -1kori +1kb DTT AnisoUrea H ActD Cispl Asynch. Sorbito control



Increasing cJun, cFos or a constitutive form of cJun in HeLa cells does not perturb HBO1 binding at origins

HeLa cells in their exponential phase of growth were transfected with plasmids encoding cJun, cFos or a constitutive JNK-phosphorylated form of cJun. 48 hours after transfection, cells were harvested and cross-linked with formaldehyde. HBO1 binding at MCM4 and TOP1 origins was investigated by ChIP (n=3).

Β



	%G0/G1	%S	%G2/M
control	49.3	22.7	28
urea	49	24.5	26.5
anisomycin	48.7	23.1	28.2
sorbitol	48.2	23.1	28.7
DTT	51.3	23	25.7
Actinomycin	D 51	24.7	24.3

Cell cycle distribution of HeLa cells exposed to different stress agents

HeLa cells in their exponential phase of growth were exposed to stress agents (in presence of serum). Cells were stained with 20μ g/ml propidium iodide (Sigma) in presence of 50μ g/ml of RNase H for 30 minutes at room temperature in the dark and extensively washed in PBS. $1x10^6$ cells were subsequently analyzed for each condition using a FACSCalibur flow cytometer and data were processed using the CellQuest software (Becton Dickinson).

Figure S3. Related to Figure 1



DNA licensing defect in HeLa cells in hyper-osmotic condition

(A) ChIP analysis of ORC2 and MCM3 binding at MCM4 and TOP1 replication origins in HeLa cells in their exponential phase of growth treated with 300mM sorbitol for 1 hour (n=3).

(B) Western blot analysis of DNA licensing factors expression and chromatin loading in HeLa cells treated with sorbitol or left untreated.

Figure S4. Related to Figure 2



Cdt1 interaction with PCNA and Skp2 is not promoted by hyper-osmotic shock

Western blot analysis of HA-Cdt1 immunoprecipitates in control and sorbitol treated cells. Nuclear extracts from HeLa cells expressing HA-Cdt1 and treated with sorbitol or not were incubated with anti-HA resin. After extensive wash, Skp2 and PCNA presence in the immunoprecipitates was tested by Western blot using specific antibodies.

Figure S5. Related to Figure 3



Efficient inhibition of ATM, p38 and JNK activity in cells treated with specific inhibitors prior to sorbitol treatment

HeLa cells in their exponential phase of growth were treated with ATM, JNK and p38 inhibitors, 30 minutes, prior to exposure to 300mM sorbitol for 1 hour. (A) ATM activity was monitored in cells by analysis of its phosphorylation level after immuno-precipitation.

(**B**) p38 activity was assessed by quantifying the phosphorylation of its substrate CREB (lordanov et al., 1997).

(C) JNK activity following SP600125 treatment was assessed by quantifying the phosphorylation of its substrate GST-cJun(Nter) in an IP-Kinase assay.

(D) JNK activity following JNK Inhibitor I treatment was monitored in cells by analysis of its phosphorylation level after immuno-precipitation.

The panels are representative of cell samples used in Figure 3A for chromatin immunoprecipitation analysis.

Figure S6. Related to Figure 3



Effect of Cdk2 kinase and p53 transcription factor on HBO1 binding at MCM4 origin

(A) HeLa cells in their exponential phase of growth were treated with a Cdk2 kinase specific inhibitor (sc-221409; Santa Cruz), prior to exposure to sorbitol. HBO1 binding was analyzed by ChIP-qPCR at the MCM4 origin (n=3).

(B) HCT116 cells, p53 positive, in their exponential phase of growth were treated with a JNK or p38 kinase specific inhibitor prior to exposure to sorbitol. HBO1 binding was analyzed by ChIP-qPCR at the MCM4 origin (n=3).

Figure S7. Related to Figure 5

Α

1 TP SP []] 29 31	SV TP 73 82	S 1 1 31	SP SP SP 8 372 394	546
В				
MEQR RVTDFF	ARRRPGPPR	APPK LACRTP	Sparpalr AP	ASATSGSRKR
ARPPAAPGRD	QARPPARR RL	RLSVDEVSSP	STPEAPDIPA	CPSPGQKIKK
STPAAGQPPH	LTSAQDQDTI	SELASCLQRA	R elgarvr al	KASAQDAGES
CTPEAEGR PE	EPCGEKAPAY	QRFHALAQPG	LPGLVLPYKY	QVLAEMFRSM
DTIVGMLHNR	SETPTFAKVQ	RGVQDMMRRR	FEECNVGQIK	TVYPASYRFR
QERSVPTFKD	GTRRSDYQLT	IEPLLEQEAD	GAAPQLTASR	LLQRRQIFSQ
KLVEHVKEHH	KAFLASL <mark>S</mark> PA	MVVPEDQLTR	WHPRFNVDEV	PDIEPAALPQ
PPATEK LTTA	QEVLARARNL	ISPRMEKALS	QLALRSAAPS	SPGSPR PALP
ATPPATPPAA	SPSALKGVSQ	DLLER IRAKE	AQKQLAQMTR	CPEQEQRLQR
LERLPELARV	LRSVFVSERK	PALSMEVACA	RMVGSCCTIM	SPGEMEKHLL
LLSELLPDWL	SLHR irtdty	VKLDKAADLA	HITARLAHQT	RAEEGL
С				
MEQR RVTDFF	ARRR PGPPRI	APPK LACRTP	SPARPALRAP	ASATSGSRKR
ARPPAAPGRD	QARPPARRRL	RLSVDEVSSP	STPEAPDIPA	CP <mark>S</mark> PGQKIKK
STPAAGQPPH	LTSAQDQDTI	SELASCLQRA	R elgarvr al	KASAQDAGES
CTPEAEGR PE	EPCGEKAPAY	QRFHALAQPG	LPGLVLPYKY	QVLAEMFR SM
DTIVGMLHNR	SETPTFAKVQ	RGVQDMMRRR	FEECNVGQIK	TVYPASYRFR
QERSVPTFKD	GTRRSDYQLT	IEPLLEQEAD	GAAPQLTASR	LLQRRQIFSQ
KLVEHVKEHH	K aflasl<mark>s</mark>pa	MVVPEDQLTR	WHPRFNVDEV	PDIEPAALPQ
PPATEKLTTA	QEVLARARNL	ISPRMEK ALS	QLALRSAAPS	SPGSPR PALP
ATPPATPPAA	SPSALKGVSQ	DLLER IRAKE	AQKQLAQMTR	CPEQEQRLQR
LERLPELARV	LRSVFVSERK	PALSMEVACA	RMVGSCCTIM	SPGEMEKHLL
LLSELLPDWL	SLHR irtdty	VKLDKAADLA	HITARLAHOT	RAEEGL

Threonine 29 is a JNK- and stress-regulated phosphorylation site

(A) Summary of previously described phosphorylation sites onto Cdt1 (Takeda et al., 2008; Chen et al., 2009; Beausoleil et al., 2004; Dephourne et al., 2008). The following residues of each phosphorylation site is also indicated. Single bars represent additional SP and TP motif, matching the JNK consensus target site.

(B) Phosphopeptide analysis of His-Cdt1 incubated with JNK1 in vitro. His-Cdt1 incubated with bulk IP (IgG) was used as a control for non-phospho-sites. Residues recovered during the mass-spectrometry analysis in both samples are indicated in bold and the phospho-sites underlined and in red.

(C) Phosphopeptide analysis of Flag-Cdt1 immunoprecipitated from HeLa cells incubated with sorbitol, sorbitol and JNK inhibitor I or left untreated. Residues recovered during the mass-spectrometry analysis in the three samples are indicated in bold. Phospho-sites are underlined and JNK-dependent phospho-site appear in red.